

Muscle NAD⁺ depletion and Serpina3n as molecular determinants of murine cancer cachexia—the effects of blocking myostatin and activins



J.J. Hulmi^{1,2,*}, F. Penna^{3,7}, N. Pöllänen^{4,7}, T.A. Nissinen¹, J. Hentilä¹, L. Euro⁵, J.H. Lautaoja¹, R. Ballaró³, R. Soliymani⁶, M. Baumann⁶, O. Ritvos², E. Pirinen^{4,8}, M. Lalowski^{6,8}

ABSTRACT

Objective: Cancer cachexia and muscle loss are associated with increased morbidity and mortality. In preclinical animal models, blocking activin receptor (ACVR) ligands has improved survival and prevented muscle wasting in cancer cachexia without an effect on tumour growth. However, the underlying mechanisms are poorly understood. This study aimed to identify cancer cachexia and soluble ACVR (sACVR) administration-evoked changes in muscle proteome.

Methods: Healthy and C26 tumour-bearing (TB) mice were treated with recombinant sACVR. The sACVR or PBS control were administered either prior to the tumour formation or by continued administration before and after tumour formation. Muscles were analysed by quantitative proteomics with further examination of mitochondria and nicotinamide adenine dinucleotide (NAD⁺) metabolism. To complement the first prophylactic experiment, sACVR (or PBS) was injected as a treatment after tumour cell inoculation.

Results: Muscle proteomics in TB cachectic mice revealed downregulated signatures for mitochondrial oxidative phosphorylation (OXPHOS) and increased acute phase response (APR). These were accompanied by muscle NAD⁺ deficiency, alterations in NAD⁺ biosynthesis including downregulation of nicotinamide riboside kinase 2 (*Nrk2*), and decreased muscle protein synthesis. The disturbances in NAD⁺ metabolism and protein synthesis were rescued by treatment with sACVR. Across the whole proteome and APR, in particular, Serpina3n represented the most upregulated protein and the strongest predictor of cachexia. However, the increase in Serpina3n expression was associated with increased inflammation rather than decreased muscle mass and/or protein synthesis.

Conclusions: We present evidence implicating disturbed muscle mitochondrial OXPHOS proteome and NAD⁺ homeostasis in experimental cancer cachexia. Treatment of TB mice with a blocker of activin receptor ligands restores depleted muscle NAD⁺ and *Nrk2*, as well as decreased muscle protein synthesis. These results indicate putative new treatment therapies for cachexia and that although acute phase protein Serpina3n may serve as a predictor of cachexia, it more likely reflects a condition of elevated inflammation.

© 2020 The Author(s). Published by Elsevier GmbH. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Keywords C26; Cancer cachexia; Activin receptor; *Nrk2*; APR; OXPHOS

1. INTRODUCTION

Cancer cachexia is a multiorgan syndrome characterised by loss of body mass (BM) due to muscle and adipose tissue wasting in the majority of cancer patients [1]. Tumour-derived factors and systemic inflammation are considered the main drivers of cachexia [1]. Inflammation leads to an increase in the concentration of cytokines and is associated with dysregulated metabolism, including mitochondrial

dysfunction, which can further contribute to muscle loss [2]. Muscle loss in cancer cachexia is strongly linked with increased morbidity and mortality [3]. Moreover, in preclinical animal models, prevention of cancer cachexia improved survival without effects on tumour growth [4–6], suggesting possible causality between muscle loss prevention and improved survival. However, the underlying mechanisms are unknown.

¹Faculty of Sport and Health Sciences, NeuroMuscular Research Center, University of Jyväskylä, Jyväskylä, Finland ²Department of Physiology, Faculty of Medicine, University of Helsinki, Helsinki, Finland ³Department of Clinical and Biological Sciences, University of Turin, Turin, Italy ⁴Research Program for Clinical and Molecular Metabolism, Faculty of Medicine, University of Helsinki, Helsinki, Finland ⁵Stem Cells and Metabolism Research Program, Faculty of Medicine, University of Helsinki, Helsinki, Finland ⁶Meilahti Clinical Proteomics Core Facility, HiLIFE, Faculty of Medicine, Biochemistry and Developmental biology, University of Helsinki, Helsinki, Finland

⁷ F. Penna and N. Pöllänen contributed equally to this work.

⁸ E. Pirinen and M. Lalowski contributed equally to this work.

*Corresponding author. NeuroMuscular Research Center, Faculty of Sport and Health Sciences, University of Jyväskylä, Rautpohjankatu 8, 40014 Finland. E-mail: juha.hulmi@jyu.fi (J.J. Hulmi).

Received April 27, 2020 • Revision received June 16, 2020 • Accepted June 23, 2020 • Available online 26 June 2020

<https://doi.org/10.1016/j.molmet.2020.101046>

Abbreviations

36B4	acidic ribosomal phosphoprotein P0	NAD ⁺	nicotinamide adenine dinucleotide
ACC	Acetyl-CoA Carboxylase	NAMPT	nicotinamide phosphoribosyl transferase
ACVR2B-Fc	activin receptor type IIB	NAPRT	nicotinate phosphoribosyltransferase
AMPK	adenosine monophosphate activated protein kinase	NMNAT	nicotinamide mononucleotide adenylyl transferase
ANOVA	analysis of variance	OXPHOS	oxidative phosphorylation
APP	acute phase protein	OXPHOSome	OXPHOS proteome
APR	acute phase response	PARP	poly(ADP-ribose) polymerase
BCA	bicinchoninic acid	PCA	Principal Component Analysis
B-H	Benjamini-Hochberg correction	PGC-1 α	peroxisome proliferator-activated receptor gamma coactivator 1 alpha
BM	body mass	PPARA	peroxisome proliferator-activated receptor-alpha
C26	colon-26 carcinoma	PPARGC1A and PPARGC1B	peroxisome proliferator-activated receptor gamma coactivator 1 alpha and beta
COX	cytochrome c oxidase	PVDF	polyvinylidene difluoride
CTRL	control mice	QPR	quinolinate phosphoribosyltransferase
DEP	differentially expressed protein	ROS	reactive oxygen species
FDR	false discovery rate	Rn18s	18S ribosomal RNA
GAPDH	glyceraldehyde 3-phosphate dehydrogenase	RT-PCR	quantitative reverse transcription polymerase chain reaction
GDF11	growth and differentiation factor 11	sACVR	soluble ligand binding domain of activin receptor type IIB
DIA nano-LC-HDMS ^E	data independent acquisition nano-liquid chromatography high definition tandem mass spectrometry	sACVR/b	sACVR administration before tumour formation
DMEM	Dulbecco's Modified Eagle's Medium	sACVR/c	continued sACVR administration before and after tumour
FBS	fetal bovine serum	SDH	succinate dehydrogenase
GA	Gastrocnemius	SDS-PAGE	sodium dodecyl sulphate polyacrylamide gel electrophoresis
HPRT	hypoxanthine phosphoribosyltransferase 1	SEM	standard error of the mean
PBS	phosphate buffered saline	Serpina3n	serpin family A member 3n
IgG	immunoglobulin G	SIRT1	sirtuin-1
IL-6	interleukin 6	STAT3	signal transducer and activator of transcription 3
IL-15	interleukin 15	SUnSET	surface sensing of translation
INSR	insulin receptor	TA	tibialis anterior
i.p.	intraperitoneal	TB	tumour-bearing
MCP-1	monocyte chemoattractant protein-1	TBP	TATA-binding protein
mTORC	mechanistic target of rapamycin complex	TCA cycle	tricarboxylic acid cycle
		TDO	tryptophan 2,3-dioxygenase
		TGF- β	transforming growth factor- β

Proteins from the transforming growth factor- β (TGF- β) superfamily are cytokines, which among others are upregulated in diseases such as cancer. For instance, the expression of TGF- β superfamily proteins, activins A and B, is increased in many human [7] and murine cancer cell lines and tumours [4]. Blood activin A is also elevated in patients with cancer cachexia [8]. Activins A and B and other TGF- β family members such as myostatin and growth and differentiation factor 11 (GDF11) are negative regulators of muscle mass [9–11]; they can be blocked using various strategies such as administration of soluble ligand binding domain of their receptor, namely, activin receptor type IIB (sACVR2B-Fc, herein referred to as sACVR), which effectively increases muscle mass in mice [4,12–15], and humans [16]. Blocking ACVR ligands prolongs survival and reverses cancer cachexia in mice [4–6]. We previously demonstrated improved survival in Colon-26 carcinoma (C26) TB-mice with continued sACVR administration after tumour formation but not when it was administered only before cancer engraftment to increase muscle mass before cachexia [4]. The administration of sACVR attenuated muscle loss in limb and respiratory muscles [4] and improved antioxidant protection in muscle tissue of TB-mice [17], without alterations in tumour growth and inflammation. However, blocking ACVR-signalling did not show major effects at either the metabolome [18] or microbiome level [19]. Thus, no explanation has been provided for the improved survival in sACVR-treated mice.

We applied 3 protocols to compare the impact of blocking ACVR2 ligands: i) administration before the tumour formation to increase muscle size only before the onset of cachexia, ii) administration both before and after the onset of cachexia, or iii) treatment after the tumour is present. These comparisons were made to further understand, at the muscle level, why improved survival was achieved when anti-cachexia treatment was continued after tumour formation but not when it was administered only as a preventive measure [4]. We first conducted muscle proteomic profiling from mice bearing the C26 cancer with or without administration of sACVR. Thereafter, we functionally validated our results to reveal Serpina3n as a predictor of cachexia, but not a generic marker of alterations in muscle mass, and discovered altered nicotinamide adenine dinucleotide (NAD)⁺ metabolism in cancer, which was rescued upon treatment with sACVR.

2. MATERIALS AND METHODS

2.1. Mice

Male BALB/c (BALB/cAnNCr) mice aged 5–6 weeks (Charles River Laboratories) were housed under standard conditions (temperature 22 °C, 12:12 h light/dark cycle) with free access to food pellets (R36; 4% fat, 55.7% carbohydrate, 18.5% protein, 3 kcal/g, Labfor, Stockholm Sweden) and water.

The treatment of animals was in strict accordance with the European Convention for the protection of vertebrate animals used for

protein mass - 750 kDa; minimum fragment ions matches per protein ≥ 7 ; minimum fragment ions matches per peptide ≥ 3 ; minimum unique peptide matches per protein ≥ 1 ; digest enzyme-trypsin; missed cleavages allowed - 2; fixed modification - carbamidomethylation C; variable modifications - deamidation (N/Q) and oxidation of methionine (M); and false discovery rate, FDR $< 4\%$. The quality of the proteomic data was evaluated by PCA and by drawing a heatmap (Heatmapper; www.heatmapper.ca) [25] of all of the quantified proteins confirming the condition-specific clustering of the data according to the study groups.

2.9. Database mining of proteomics data

Relative protein quantification between samples using precursor ion intensities was performed with Progenesis Q1™ Informatics for Proteomics software (Version 3.0, Nonlinear Dynamics/Waters). To identify differentially expressed proteins (DEPs), *P* values (by ANOVA) and fold change (FC) were set to < 0.05 and $> |1.3|$, respectively. The quantification was performed based on ≥ 2 unique peptides. The filtered, false discovery rate (FDR < 0.05) corrected lists served as inputs into Ingenuity Pathway Analysis (IPA, Ingenuity Systems, Redwood City, CA; www.ingenuity.com).

2.10. Protein extraction and Western blotting

TA muscle ($n = 7-8$ per group in experiment 1 and $n = 5-6$ in experiment 2) was homogenised in ice-cold buffer with proper protease and phosphatase inhibitors [4,12] and centrifuged at 10 000 g for 10 min at $+4^\circ\text{C}$ except for the SUnSET protein synthesis measurements, where samples were centrifuged at 500 g for 5 min. Total protein content was determined by using the bicinchoninic acid protein assay (Pierce, Thermo Scientific) with an automated KoneLab device (Thermo Scientific). Muscle homogenates containing 30 μg of protein were solubilised in Laemmli sample buffer and heated at 95°C to denature proteins, separated by SDS-PAGE, and transferred to PVDF membrane, followed by overnight probing with primary antibodies at $+4^\circ\text{C}$. Muscle proteins were visualised by enhanced chemiluminescence using a ChemiDoc XRS or ChemiDoc MP device (Bio-Rad Laboratories) and quantified with Quantity One software version 4.6.3 (Bio-Rad Laboratories, Hercules, California, USA) or with Image Lab software (version 6.0; Bio-Rad Laboratories), respectively. In the case of puromycin-incorporated proteins, the intensity of the whole lane was quantified. Ponceau S staining and glyceraldehyde 3-phosphate dehydrogenase (GAPDH, ab9485: Abcam, Cambridge, USA) were used as loading controls, and the muscle protein level results were normalised to the mean of Ponceau S and GAPDH (experiment 1) or to the overall intensity on stain-free gels (experiment 2). To assess serum proteins, samples ($n = 7-9$ per group) were diluted 1:4, and the results were normalised to the total protein intensity on the membrane by using stain-free gels. Primary antibodies were against serpin family A member 3n (Serpina3n, AF4709: R&D Systems, Minneapolis, USA), p-STAT3 at Tyr⁷⁰⁵, total STAT3, p-ACT at Ser⁷⁹, p-Akt at Ser⁴⁷³ and total Akt (Cell Signalling Technology, Danvers, USA), and fibrinogen (Abcam ab27913, Cambridge, UK).

2.11. RNA extraction, cDNA synthesis, and quantitative real-time PCR

Total RNA was extracted from gastrocnemius samples ($n = 7-8$ per group) by using QIAzol and purified with the RNeasy Universal Plus kit (Qiagen) according to manufacturer's instructions, resulting in high-quality RNA. RNA was reverse transcribed to complementary DNA (cDNA) with the iScript™ Advanced cDNA Synthesis Kit (Bio-Rad Laboratories) according to kit instructions. Real-time qPCR was

performed in triplicate according to standard procedures by using iQ SYBR Supermix (Bio-Rad Laboratories) or Maxima SYBR Green qPCR Master Mix (2x; ThermoFisher Scientific) with CFX96 or CFX384 Touch™ Real-Time PCR Detection Systems and CFX Manager software (Bio-Rad Laboratories), respectively. Data analysis was conducted by using the efficiency corrected $\Delta\Delta\text{Ct}$ method. Based on the lowest variation between and within the groups of the potential housekeeping genes (*Rn18S*, *Gapdh*, *36b4*, *Hprt* or *Tbp*), *36b4* or the mean of *36b4* and *Hprt* was chosen. Primers used are listed in Supplementary methods (Table S1).

2.12. NAD⁺ metabolism and mitochondrial *in situ* analyses

NAD⁺ and NADH were measured from gastrocnemius muscle ($n = 6$ per group) by using slightly modified conventional colorimetric enzymatic assays (see NADmed™ <https://www.helsinki.fi/en/researchgroups/mitochondrial-medicine/nadmed>). The activity of main cellular NAD⁺ consumers, poly (ADP-ribose) polymerases (PARPs) was determined from gastrocnemius muscle with the HT Colorimetric PARP/Apoptosis Assay Kit (Trevigen, Inc., Gaithersburg, MD, USA) according to manufacturer's instructions ($n = 5-6$ per group).

For mitochondrial activity *in situ* analysis, serial cross-sections (8 μm) from TA muscle were cut on a cryomicrotome (at -25°C ; experiment 1, encompassing mice from CTRL, C26 + PBS and C26 + sACVR/c groups). Mitochondrial activity was investigated by assessing the *in situ* activity of succinate dehydrogenase (SDH, complex II) and cytochrome c oxidase (COX, complex IV) from frozen TA muscle sections by using histochemistry. Cross-sectional images were digitally captured by using a Zeiss Axioplan 2 light microscope (Carl Zeiss AG, Germany) with a ProgRes C7 camera and ProgRes CapturePro 2.9.0.1 software (Jenoptik AG, Germany). Two fields-of-view per muscle cross-section were captured with a $5\times$ magnification. A sample was excluded if cutting of the cross-section was unsuccessful (resulted in $n = 5-7$ per group). The COX and SDH *in situ* activities were analysed with Fiji/ImageJ (NIH, Bethesda, MD, USA) plugin Trainable Weka Segmentation [26]. Image resolution was reduced to 1/4 of the original because of the computational power required for analysis. Regions of interest were determined separately for each image and, after splitting channels, applied to green channel images to obtain optical density values of different fibre types. Optical density was assessed from all fibres and from high COX and high SDH fibres. Muscle distribution of mitochondria-rich, high SDH and COX fibres was calculated by using the ImageJ plugin Cell Counter and represented as percentage of high optical density fibres from all fibres.

2.13. Flow cytometry analysis to characterise splenomegaly in cancer

In the second experiment, the spleens of mice derived from 3 experimental groups ($n = 6-8$ per group) were harvested under sterile conditions, and single cell suspensions were prepared by gently pressing the minced spleens through 70 μm cell strainers. The total number of cells was calculated by counting the cell concentration by using a Beckton Dickinson Accuri C6 flow cytometer. Ten⁶ cells were then incubated with 0.4 mg/ml RNase A for 30' and stained with 18 $\mu\text{g}/\text{ml}$ propidium iodide to distinguish between erythrocytes and nucleated cells and assess the percentage of proliferating cells.

2.14. Statistical analysis

Analyses (for proteomics analyses details, see above) were conducted with a general linear model ANOVA or Kruskal–Wallis test and then a two-tailed unpaired Student's *t* test or Mann–Whitney U test, respectively, when appropriate. Planned comparisons in all analyses

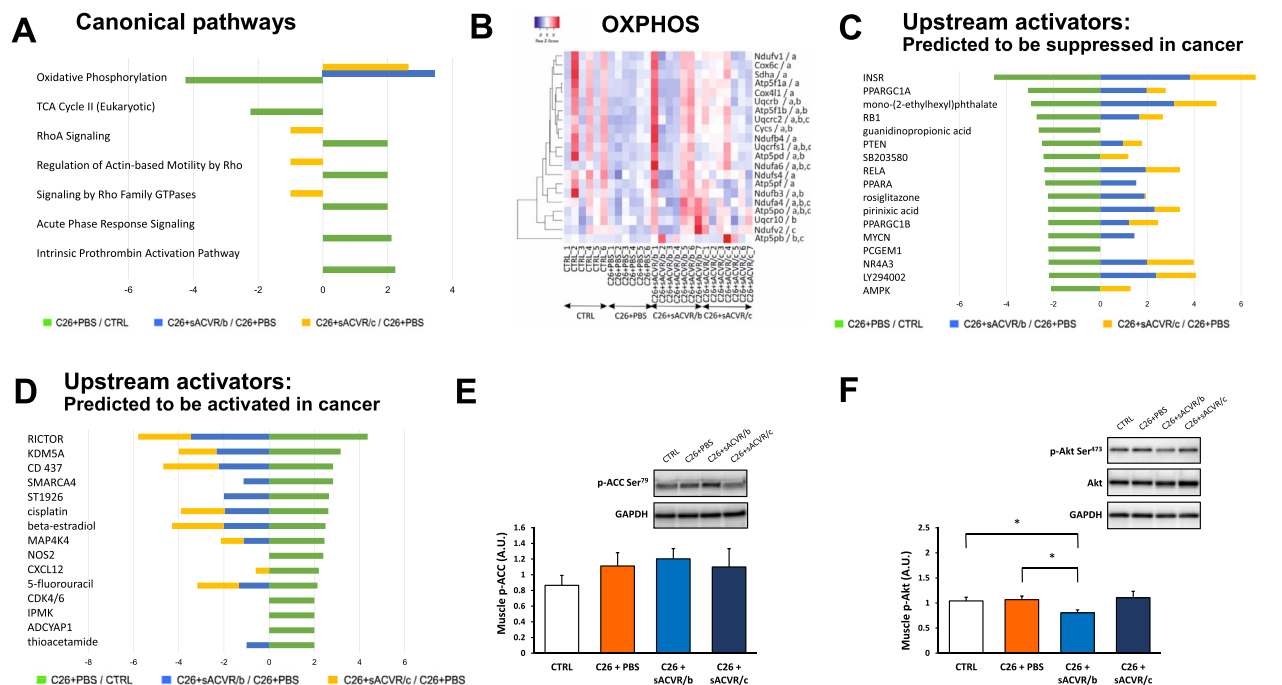


Figure 2: Ingenuity pathway analysis reveals pathways and upstream regulators affected in muscle by cancer and anti-cachexia treatment in experiment 1. Canonical pathway analysis (A). Pathways with significant z-scores ($>|2|$) in one of 3 comparisons are presented, and if available, z-scores of those pathways are also shown from other comparisons. Heat map representation (B) of OXPHOS proteins with largest differential changes in their expression, in 3 comparisons: (a) C26 + PBS vs CTRL, (b) C26 + sACVR/b vs C26 + PBS, and (c) C26 + sACVR/c vs C26 + PBS. The criteria used for assessment were as follows: fold change, $FC > |1.3|$, FDR corrected p value < 0.05 , and ≥ 2 unique peptides used for quantitation. An expression-based heat map using average linkage and the Spearman rank correlation distance measures. Upstream analysis pinpointing upstream regulators predicted to be suppressed (C) or activated (D) in C26 + PBS vs CTRL comparison (z-score $> |2|$) and if available, z-scores of those pathways are also shown from other comparisons. Phosphorylated ACC at ser⁷⁹ as a proxy for AMPK activation (p-ACC; E) and p-Akt at Ser⁴⁷³ as a proxy for mTORC2 activation (F). * = $P < 0.05$ using a two-tailed unpaired Student's t test.

were CTRL vs C26 + PBS and within TB-mice, either sACVR/c or sACVR/b (or sACVR in the second experiment) vs C26 + PBS and sACVR/c vs sACVR/b. When no clear effect of the sACVR administration was observed, the C26 cancer effect was analysed by pooling C26 groups vs CTRL to simplify the results and the figures. Pearson's correlation coefficient was used to assess the correlations. The level of significance was set at $P < 0.05$. Data were expressed as means \pm SEM if not otherwise mentioned. Statistical analyses were performed with IBM SPSS Statistics version 24 for Windows (SPSS, Chicago, IL) and with GraphPad Prism 8.0.

3. RESULTS

3.1. Characterisation of muscle proteome

Previously, we demonstrated that C26 cell inoculation resulted in muscle and fat wasting. Treating these TB-mice with sACVR before and after tumour formation prevented muscle loss and prolonged survival without affecting tumour size [4]. In this study, using label free proteomics, we identified 1232 proteins extracted from the murine muscles of these mice [4]. Notably, 876 proteins were quantified, followed by the exclusion of proteins considered contaminants, for example, blood haemoglobins or skin keratins (Table S2). The features of all groups were assessed by principal component analysis (PCA), which demonstrated a good group separation for TB and control mice (for a summary, see the panels in Figure S1). PCA also revealed 2 biological outliers that we excluded from subsequent analyses, resulting in $n = 6-7$ mice per group. We identified 119 DEPs in PBS-

treated TB-mice. Among these, 53 proteins exhibited a higher expression level in TB-mice, and 66 were downregulated. sACVR injection resulted in increased muscle size [4], accompanied by 126 DEPs (53 upregulated and 73 downregulated) with the prophylactic and continued (sACVR/c) administration, and by 117 DEPs (58 up- and 59 downregulated in their expression) in the prophylactic/discontinued (sACVR/b) administration, respectively, when compared to PBS-treated TB-mice. For the purpose of stringency, only the most altered proteins ($FC > |2|$) are presented in Figure 1, and the comparisons between the administration modalities are in Table S2.

3.2. Pathway analysis reveals decreased mitochondrial OXPHOS signature in TB-mice: a response attenuated in sACVR-treated mice

Canonical pathway analysis of DEPs revealed a signature of decreased mitochondrial oxidative phosphorylation (OXPHOS) in the PBS-treated TB-mice (Figure 2A). Eighteen DEPs within this pathway included subunits of OXPHOS complex I (NADH: ubiquinone oxidoreductase), complex II (succinate dehydrogenase, SDH), complex III (ubiquinol-cytochrome c reductase), complex IV (cytochrome c oxidase, COX), and complex V (ATP synthase), suggesting a wide-spectrum down-regulation of OXPHOS proteome (OXPHOSome; Figure 2B). This downregulation was in part (several proteins of CI-IV and ATP synthase subunits) prevented by sACVR/b (7 DEPs significantly upregulated compared to C26+PBS) and sACVR/c administration (12 DEPs upregulated vs C26+PBS), without an observable difference between sACVR/c and sACVR/b (Figure 2B). TB-mice also exhibited a consistent decrease in the expression of mitochondrial TCA cycle proteins (5 out

of 5 quantified), which is in line with the reported decline in citrate synthase activity in the same TB-mice [4]. However, the blocking of ACVR ligands had no marked effect on the TCA cycle (Figure 2A).

To further understand how the altered OXPHOS signature affects mitochondrial metabolism, we determined *in situ* activities of complex II (SDH) and complex IV (COX) from the CTRL, C26+PBS, and C26+sACVR/c groups. In line with the OXPHOS, in some results showing complex II protein subunit downregulation, the percentage of muscle fibres endowed with high SDH activity (and thus mitochondria-rich fibres) significantly decreased in the PBS-treated TB-mice, and this decrease was not significant in mice administered by continued sACVR/c (Figure 3A). Different administration protocols did not induce changes in total SDH and COX activities (Figure S2) or in the percentage of muscle fibres with high COX activity (Figure 3B). In summary, decreased *in situ* SDH activity in TB-mice validated dysregulated OXPHOS in cancer cachexia.

3.3. Decreased oxidative and anabolic signalling in TB-mice predicted by upstream factor analysis, the latter response rescued by sACVR

Unbiased upstream-regulator analysis suggested possible upstream regulators to explain the dysregulated mitochondrial signature (Figure 2C,D). More explicitly, it predicted that transcriptional regulatory proteins important for muscle mitochondrial biogenesis, OXPHOS and fatty acid oxidation, such as peroxisome proliferator-activated receptor gamma coactivator 1 alpha, and beta (PPARGC1A and PPARGC1B, namely, PGC-1 α/β) [27], peroxisome proliferator-activated receptor-alpha (PPARA, i.e., PPAR α) [28], and surprisingly, AMP-activated protein kinase (AMPK) [29] are also significantly suppressed (i.e. reduced activation, z-score < |2|) in TB-mice (Figure 2C). However, phosphorylated ACC at Serine⁷⁹ status, often used as a proxy for AMPK activation, did not confirm suppressed AMPK-signalling in cachexia (Figure 2E). Among predicted upstream regulators, PGC-1 α and PGC-1 β were rescued by sACVR (Figure 2C).

Related to the decreased muscle mass in TB-mice and its treatment by continued sACVR administration [4], the reduced activation of insulin receptor (INSR) was predicted to be strongly linked to the C26+PBS vs CTRL scenario (a consistent decrease in all 22 DEPs in the dataset for

the insulin receptor; Figure 2C). Particularly, the sACVR/c administration fully restored the activation of INSR in the same analysis (Figure 2C). The predicted upstream regulators activated in TB-mice are presented in Figure 2D. Connected to this pathway is the main muscle size regulating hub, mTORC1, which was shown to be suppressed in TB-mice and rescued by sACVR [4]. Surprisingly, Rictor, which refers to a mechanistic target of rapamycin complex 2 (mTORC2), was predicted by IPA to be strongly activated in PBS-treated TB-mice, and this response was prevented specifically in sACVR/c treated mice (Figure 2D). The prediction implies that activation of mTORC1 and mTORC2 is regulated in an inversed manner in cachexia and after the blocking of activin receptor ligands. Therefore, phosphorylated Akt at Serine⁴⁷³ status, a standard proxy for mTORC2 activation, was analysed. However, the result of this analysis did not confirm activated mTORC2-signalling in cachexia (Figure 2F).

3.4. Disturbed NAD⁺ homeostasis in TB-mice is rescued by the continued sACVR administration

NAD⁺ metabolism is crucial for mitochondrial oxidative metabolism. Namely, the NAD⁺/NADH redox couple regulates the activity of the TCA cycle and OXPHOS, and the gene expression and activity of sirtuin 1 (Sirt1), the key inducer of PGC-1 α activity [30]. Therefore, oxidised and reduced concentrations of NAD⁺ were assessed. We observed that the levels of both oxidised (NAD⁺) and reduced NAD (NADH) decreased in PBS-treated TB-mice ($P < 0.05$), and unlike sACVR/b, sACVR/c restored the depleted NAD⁺ levels; for NADH, this effect was not statistically significant (Figure 4A,B).

To uncover the mechanism behind NAD⁺ depletion, we next analysed NAD⁺ consumption and biosynthesis pathways in TB-mice. Given that the activity of the major cellular NAD⁺ consumers, PARPs, remained unchanged, the NAD⁺ depletion in TB-mice is unlikely to result from PARP-dependent NAD⁺ utilisation (Figure 4C). Notably, the expression analysis of the main muscle genes involved in NAD⁺ biosynthesis pathways (salvage and Preiss–Handler pathways) revealed a strongly diminished muscle isoform of nicotinamide riboside kinase (*Nrk*), *Nrk2*, in PBS-treated TB-mice, and sACVR/c completely rescued it ($P < 0.05$; Figure 4D). Similar to NAD⁺ levels, the sACVR/b administration modality demonstrated no effects on *Nrk2* expression (Figure 4D). By

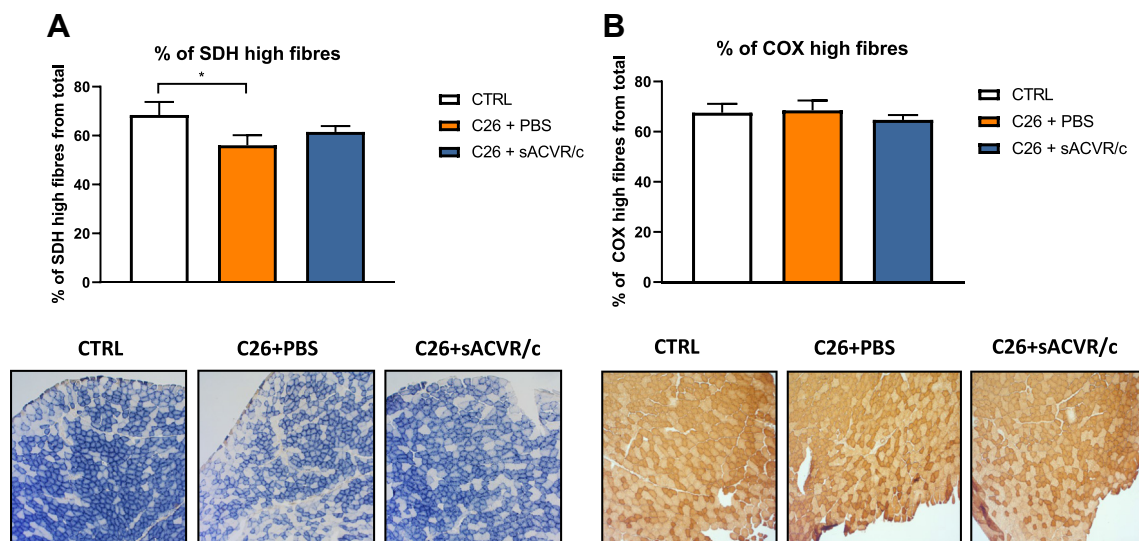


Figure 3: Muscle mitochondrial activity *in situ* is in part altered in cancer in experiment 1. Percentage of fibres with high SDH (A) or COX (B) activities. * = $P < 0.05$ using a two-tailed unpaired Student's t test.

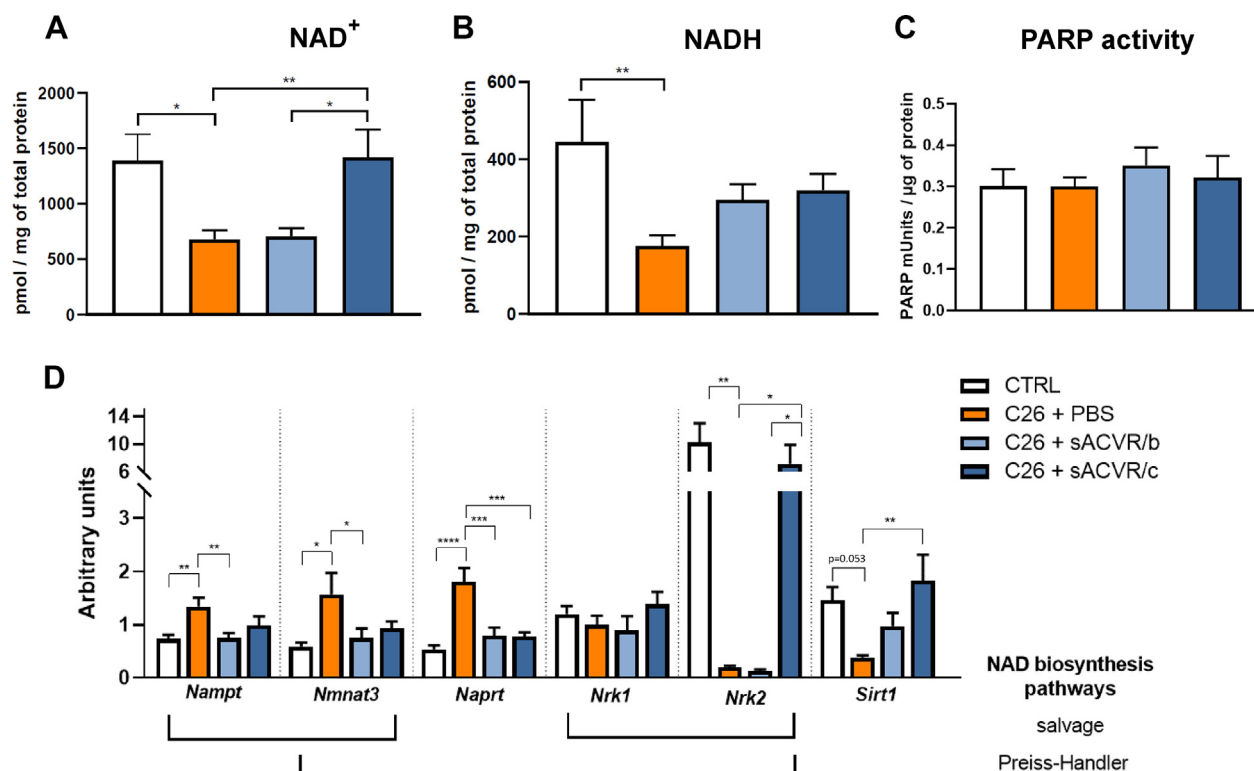


Figure 4: NAD⁺ and its regulators are affected in TB-mice and in part rescued by anti-cachexia treatment in experiment 1. Contents of NAD⁺ (A) and NADH (B), and PARP activity (C). Gene expression of the regulators of NAD⁺ biosynthesis (D). * = $P < 0.05$, ** = $P < 0.01$, and *** = $P < 0.001$ when using a two-tailed unpaired Student's t test.

contrast, the expression of the ubiquitously expressed isoform, *Nrk1*, was similar in all groups (Figure 4D). The expression of other enzymes of the NAD⁺ biosynthesis pathway, namely, nicotinamide phosphoribosyltransferase (*Nampt*), nicotinamide adenyl transferase 3 (*Nmnat3*), and nicotinate phosphoribosyltransferase (*Naprt*) increased in PBS-treated TB-mice, a feature that was normalised by sACVR (Figure 4D).

The *de novo* NAD⁺ biosynthesis pathway is considered to operate at a low level in skeletal muscle and thus may have low physiological significance. Nevertheless, we observed increased tryptophan 2,3-dioxygenase (*Tdo*) and quinolinate phosphoribosyltransferase (*Qprt*) expression in the PBS-treated TB-mice, and these responses tended to be preserved in the sACVR-treated condition (Figure S3). Downstream to NAD⁺, TB-mice demonstrated downregulation of NAD⁺ dependent *Sirt1* ($P = 0.053$, Figure 4D), which acts as an important protein deacetylase, protecting cells from metabolic stress. In line with the NAD⁺ and *Nrk2* results, *Sirt1* expression was rescued by sACVR/c, and less so by sACVR/b (Figure 4D). Overall, our data suggest that cancer cachexia is characterised by muscle NAD⁺ deficiency which can be restored by sACVR when administered before and after tumour injection.

3.5. Therapeutic ACVR ligand blockade restores muscle mass, protein synthesis, NAD⁺ levels and *Nrk2*

In addition to the prophylactic and/or prophylactic and continued ACVR blocking, we next conducted a more translatable experiment in which the sACVR administration was initiated after tumour formation. We observed that sACVR treatment increased BM (Figure 5A) and rescued the decrease in muscle mass (Figure 5B) and protein synthesis in TB-mice (Figure 5C) without affecting the tumour growth (Figure 5B). We also replicated the findings of experiment 1 and showed that muscle

NAD⁺ is depleted in C26 TB-mice even when cachexia is very mild (Figure 5D), and this was accompanied by decreased *Nrk2* expression ($P = 0.052$; Figure 5E). Therapeutic sACVR treatment restored levels of NAD⁺ (Figure 5D), and a similar trend was observed for *Nrk2* ($P = 0.095$; Figure 5E); both results are in line with the results of sACVR/c administration. To support the findings from experiment 1, we also replicated the results of *Nmnat3* and *Naprt* gene expression but not of *Nampt* (Figure 5E). However, the expression of *Nrk1* and *Sirt1* was very low and thus not quantified. In summary, these data validated the tumour-effects observed in a previous experiment and showed that muscle NAD⁺ deficiency can be restored by exposure to therapeutic sACVR.

3.6. Pathway and upstream analyses reveal acute phase response in TB-mice

Pathway analysis predicted involvement and upregulation of acute phase proteins (APPs) reflecting acute phase response (APR) in cancer (Figures 2A and 6A). Corresponding upstream analysis indicated that activated STAT3 ($P = 2.17 \times 10^{-7}$, Z-score 1.45, not shown) may represent a regulator for this response, as shown by Bonetto et al. [31]. Subsequently, Western blotting analysis confirmed increased phosphorylation of STAT3 in TB-mice (Figure 6B). The STAT3 signalling pathway is downstream to IL-6 and MCP-1, both being elevated in the serum of TB-mice [4], which may connect mitochondrial dysfunction to muscle cachexia [2].

3.7. APP Serpina3n, the most altered protein in TB-mice, is associated with the loss of BM but not affected by the anti-cachexia treatment

Among identified DEPs within the APR pathway, Serpina3n (human orthologue of Serpina3, also known as Alpha-1-antichymotrypsin)

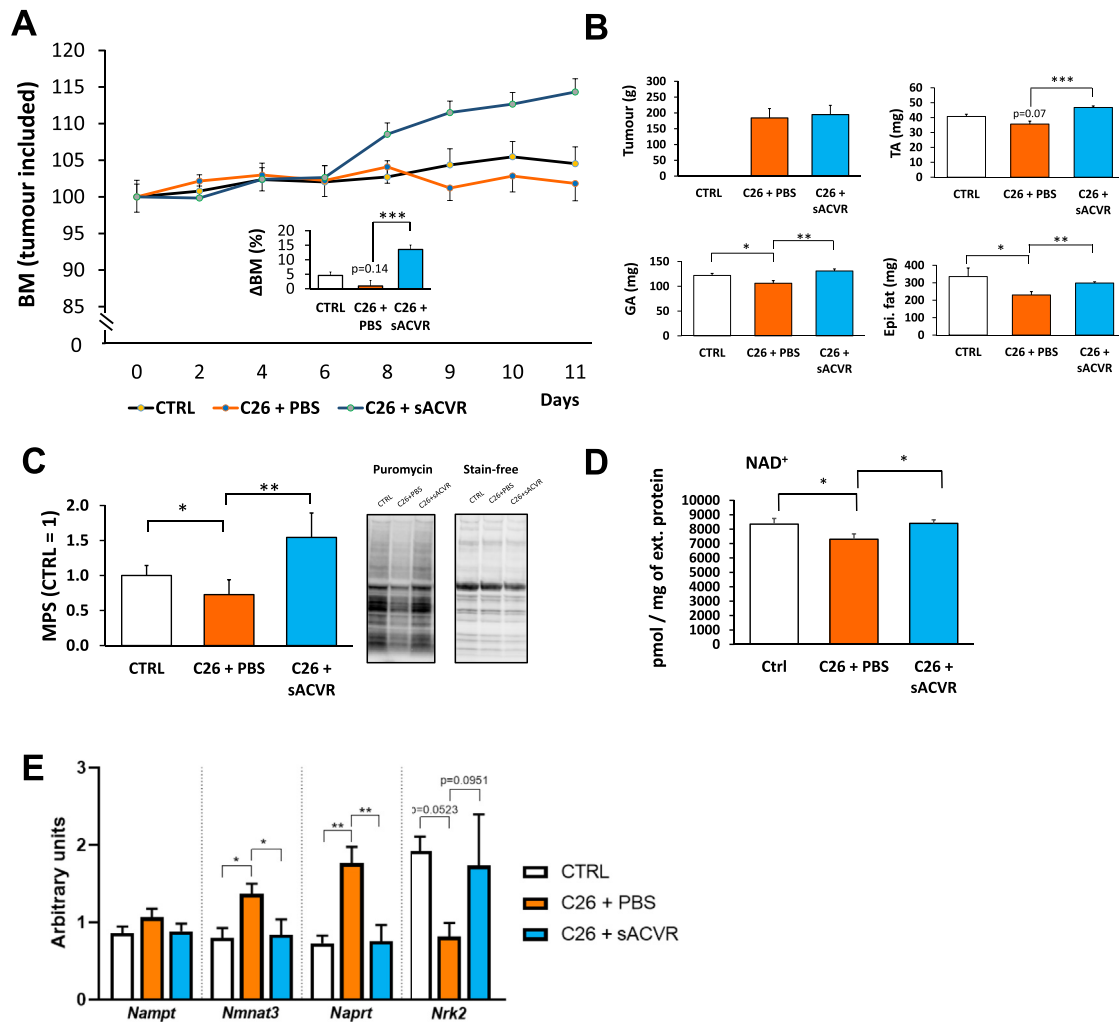


Figure 5: Anti-cachexia treatment after tumour inoculation (experiment 2) reveals increased body mass, muscle masses, and muscle protein synthesis and replenished NAD⁺. Changes in body mass (BM; A), tissue masses (B), muscle protein synthesis (C), contents of NAD⁺ (D), and gene expression of the regulators of NAD⁺ biosynthesis (E) in control mice and in mice treated with sACVR or PBS. Epi. fat = epididymal fat pad. * = $P < 0.05$, ** = $P < 0.01$, and *** = $P < 0.001$ when using a two-tailed unpaired Student's t test.

represented the most strongly upregulated protein (Figures 1B and 6A), whose increased expression was validated in cachectic muscle by Western blotting (Figure 6C). Elevated Serpina3n levels were also observed in the serum of TB-mice (Figure 6D). The increase in Serpina3n protein was probably due to the transcriptional regulation of the gene because the level of muscle mRNA expression of *Serpina3n* increased in TB-mice (Figure S4), and it correlated well with the Serpina3n protein level ($r = 0.67$, $P = 0.001$). Serpina3n was higher in the sACVR/b group than in the sACVR/c ($P < 0.001$) group, without an observable difference when compared with PBS-treated TB-mice (Figure 6A,C). A similar effect was traced on the qPCR analysis (Figure S4), demonstrating that Serpina3n protein and mRNA levels are most upregulated in a group of mice with the strongest decrease in BM during the last days before the tissue collection [4]. These results and the earlier evidence [32] suggest that Serpina3n may represent a biomarker related to changes in muscle/BM. Indeed, muscle Serpina3n protein strongly correlated with the loss of BM at 9–11 days after tumour cell engraftment ($r = -0.77$, $P < 0.001$) in TB-mice (Figure 5E). A similar observation was made at the mRNA level

($r = -0.84$, $P < 0.001$, Figure S5A). At this time point, the loss of BM predicted survival (Cox regression: $B = 0.90$, $P < 0.001$) in another group of mice [4]. The proteomics result was validated by Western blotting in TB-mice ($r = -0.64$, $P < 0.001$, Figure 6E). In serum ($r = -0.69$, $P < 0.001$) and in the liver ($r = -0.82$, $P < 0.001$), Serpina3n levels also correlated with BM loss at the same time point (Figure S5A). These results demonstrate that high Serpina3n expression (at the mRNA and protein level) is related to a significant loss of BM in TB-mice.

The gene expression of other APPs, namely, *Saa1* and *Fibrinogen G* (Figure S4), as well as serum Fibrinogen, were elevated in TB-mice (Figure S5B). Unlike Serpina3n, serum Fibrinogen correlated with BM loss only when all mice were included ($r = -0.58$, $P < 0.001$), but not in TB-mice ($r = -0.32$, $P = 0.154$; Figure S5B). In skeletal muscle, APP components were not affected by sACVR (Figure 6A). We subsequently conducted a correlation analysis including all proteins identified in muscle proteomics. Among those, Serpina3n clearly correlated most strongly with the loss of BM at 9–11 days after tumour cell engraftment followed by fibrinogen G (data not shown).

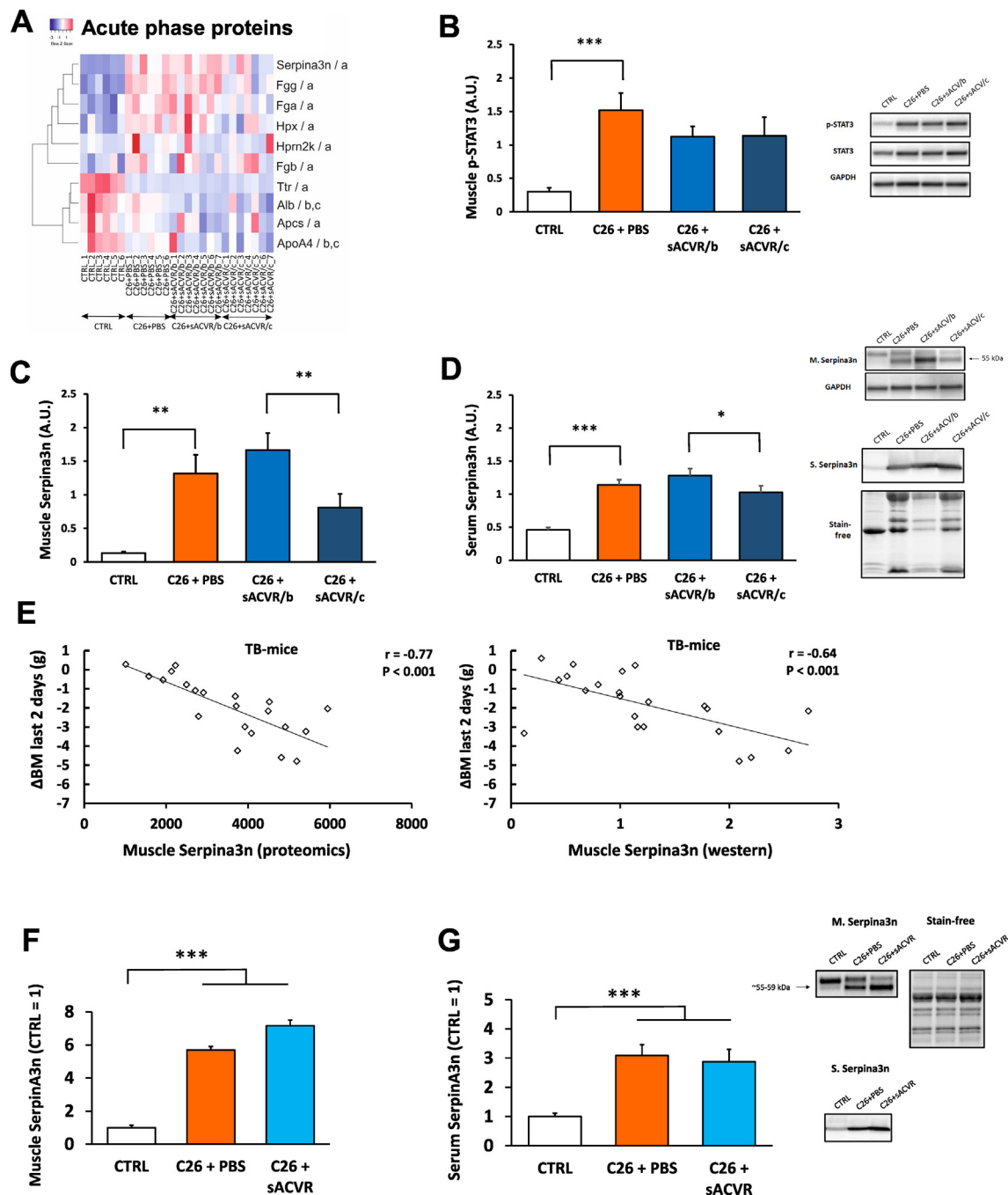


Figure 6: APR is activated in TB-mice independent of anti-cachexia treatment. Heat map representation of differential proteome changes of APPs in experiment 1. The analyses in 3 conditions with the highest fold change ratio are shown ($FC > 1.3$, FDR corrected P value < 0.05 and ≥ 2 unique peptides used for quantitation), namely: (a) C26 + PBS vs control, (b) C26 + sACVR/b vs C26 + PBS, and (c) C26 + sACVR/c vs C26 + PBS. An expression-based heat map is using average linkage and the Spearman rank correlation distance measures (A). Phosphorylated STAT3 (p-STAT3; B) and Serpina3n quantifications in muscle (C) and in serum (D). Correlation analysis of Serpina3n in the muscle, quantified either by label free proteomics or with Western blotting and cachexia (a decrease in the body mass, BM; E). Serpina3n in muscle (F) and in serum (G) in mice treated with sACVR/PBS after tumour cell inoculation in experiment 2. * = $P < 0.05$, ** = $P < 0.01$, and *** = $P < 0.001$ when using a two-tailed unpaired Student's t test.

3.8. Cachexia-associated Serpina3n reflects the inflammatory state

In the second experiment, we replicated the findings of increased muscle (Figure 6F) and serum (Figure 6G) Serpina3n independently of sACVR treatment, even though sACVR rescued cachexia and muscle protein synthesis. Furthermore, muscle Serpina3n correlated with BM loss in TB-mice only when sACVR-treated mice (with strongly

increased BM, muscle mass, and muscle protein synthesis) were excluded from the analysis (Figure S6). To further analyse the impact of inflammation in TB-mice and whether Serpina3n reflects on it, we analysed the spleen size and its morphology. Spleen size increased in TB-mice, and this was not affected by the sACVR treatment (Figure S7). To investigate the C26-induced spleen enlargement, namely, splenomegaly in more detail, a flow cytometry analysis was

conducted, which demonstrated the process that stemmed from both increased erythrocyte engulfment and white blood cell proliferation (Figure S7). Spleen mass, as well as serum IL-6, correlated with muscle Serpina3n (Figure S6).

4. DISCUSSION

In this study we characterised the muscle proteome of TB-mice with or without anti-cachexia treatment. Our proteomics results demonstrated a reduction in the expression of proteins implicated in OXPHOSome, a finding further corroborated by measurements pinpointing dysregulated NAD⁺ metabolism and alterations in NAD⁺ biosynthesis including downregulation of *Nrk2* in cancer cachexia. These changes were rescued by sACVR when administered to TB-mice. In subsequent experiments we demonstrated that sACVR treatment that commenced after tumour formation prevented a cancer-induced decrease in muscle protein synthesis and NAD⁺ but not splenomegaly. Finally, we established evidence for acute phase protein Serpina3n as the most consistent predictor of untreated cachexia in murine muscle proteome but not as a generic marker of alterations in muscle mass.

The decrease in the OXPHOSome, the lowered expression of components in the TCA-pathway, and the low percentage of muscle fibres with high SDH activity in TB-mice support the published observations of mitochondrial dysfunction in cancer cachexia demonstrated by using proteomics [33], transcriptomics [34], and mitochondrial oxidative enzyme activity analyses [4,35]. However, based on Western blot assay, the protein content of few OXPHOS proteins has also been reported to remain relatively unaltered in cachectic TA muscle [4]. In cancer cachexia, muscle mitochondrial dysfunction may occur in part through cytokine-signalling such as IL-6 — STAT3 axis [2], confirmed by this study; alternatively or complementary to this, reduced physical activity observed in these TB-mice [4] may contribute to the process of dysregulated mitochondria [2] that per se may be sufficient to induce muscle atrophy [36]. Utilising proteomics, we demonstrated that blocking ACVR ligands attenuated these changes in OXPHOSome, but no significant differences were observed in the response between the administration modalities. Previously, blocking ACVR ligands has either had no effect [15] or even diminished mitochondrial oxidative capacity [14,37,38], and decreased OXPHOS-related protein expression in muscle [39] while inducing fast and glycolytic transformation [40]. However, blocking ACVR ligands has also resulted in positive outcomes in various disease, injury, and ageing models [4–6,15,40–42]. Our results imply that blocking ACVR ligands can have context-dependent effects on muscle oxidative metabolism. Whether the rescue of OXPHOSome is linked to improved survival [4] requires further research.

As predicted by the upstream-regulator analysis, the diminished activities of PGC-1 α/β and PPAR α may contribute to the global OXPHOSome alterations in cachectic muscle. Their deletion causes impairment of mitochondrial OXPHOS transcription and oxidative metabolism especially in stress conditions in mice [27,28]. We have observed a decrease in muscle PGC-1 α in mice inoculated with C26 cells [35], and a similar tendency was observed in these mice, in TA muscle [4]. Moreover, dysregulated NAD⁺/sirtuin metabolism may contribute to the altered OXPHOSome signature in cachectic muscle. Notably, both oxidised and reduced NAD⁺ levels were significantly lowered in untreated TB-mice. In addition, a moderate decrease in the key regulator of muscle mitochondrial oxidative metabolism and stress defence, NAD⁺-dependent *Sirt1*, was observed in cancer cachexia, as was shown in TB-mice in relation to the severity of cancer cachexia [43]. The continued and post-tumour ACVR treatment replenished NAD⁺ levels, and continued administration also normalised the expression of *Sirt1*

similar to the predicted activities of its downstream targets PGC-1 α/β . NAD⁺ depletion and a subsequent disturbance in cellular redox status can alter the function of various pathways such as mitochondrial energy production, cellular signalling, chromatin modulation, DNA repair, ROS defence, inflammation and immune responses, and circadian rhythm [44]. Previously, we demonstrated that sACVR altered redox balance and offered antioxidant protection in the muscles of these TB-mice by restoring the decreased levels of reduced glutathione (GSH) [17]. Therefore, sACVR can enhance the function of a wide range of cellular processes. In line with our results, several animal studies have demonstrated that NAD⁺ boosting is a promising therapy option to treat diseases with multifactorial pathology, for example, muscle dystrophy and myopathy, obesity, diabetes, fatty liver, and cardiac diseases [44]. Further studies should investigate whether NAD⁺ boosters offer an advantage in cancer cachexia. Overall, our work establishes a novel finding of NAD⁺ depletion in cancer cachexia, which can be restored by sACVR treatment in TB-mice.

This study also revealed that NAD⁺ depletion in cachectic muscle is associated with the altered expression of genes involved in NAD⁺ biosynthesis pathways but not with enzymatic activities of NAD⁺ consumers. Namely, the decline in NAD⁺ levels probably did not originate from the increased activity of PARPs, the major NAD⁺ consumers in cells [45], because the total PARP activity was unaltered in TB-mice. The analysis of NAD⁺ biosynthesis proposes *Nrk2* (*Nmrk2*), an enzyme involved in the salvage pathway, is potentially associated with low muscle NAD⁺ content in cancer cachexia. By contrast, the gene expression of several other enzymes in the NAD⁺ biosynthesis pathway was elevated in PBS-treated TB-mice. These findings are puzzling because the deletion of *Nrk2* has not been demonstrated to affect muscle NAD⁺ levels because *Nrk2* is redundant with other proteins such as *Nrk1* in the NAD⁺ salvage pathway [46,47]. Thus, the question raised is whether muscle *Nrk2* downregulation results in a depletion of NAD⁺ levels in cancer cachexia or whether *Nrk2* downregulation is related to its other functions such as modulation of muscle mass. Another question raised here is whether *Nrk2* downregulation is related to its other functions such as regulation of muscle fibre size, as reported in the literature [47]. *Nrk2* deficiency has also been shown to increase muscle expression of cytokine *Il-15*, a potent regulator of muscle growth. We, however, did not observe major changes in muscle *Il-15* (Figure S3). Overall, our studies revealed downregulation of *Nrk2* in cancer cachexia and its restoration by sACVR. Nevertheless, further research should investigate the regulation of *Nrk2* via TGF- β superfamily cytokines and the role of *Nrk2* in the maintenance of muscular NAD⁺ pool during pathological conditions such as cancer cachexia. Further research should also examine whether the myostatin/activin blockage repletes NAD⁺ and *Nrk2* deficiencies via direct action or by interfering with cachexia-altered signalling events.

APR is an early response in cancer belonging to the innate immune system characterised by increased expression of positive APPs [48]. The APR protein Serpina3n belongs to a family of serine protease inhibitors. Serpina3n contains a signal peptide and is secreted into C2C12 muscle cell media after atrophic stimuli (i.e. glucocorticoids) [32], and in humans, it may be exported to blood through small vesicles [49], suggesting that Serpina3 (or Serpina3n) may indeed be a myokine. As demonstrated in proteomics experiments, Serpina3n was the most strongly associated protein with the loss of BM. Similarly, elevated Serpina3n levels in plasma and liver strongly correlated with cachexia. This occurred at a time point in which cachexia was predictive for a large variation in survival [4]. Our results are in accordance with the results of human studies connecting increased APR with compromised survival in cancer cachexia [50,51]. Although APR may

assist in limiting host infection and tissue injury, its prolonged action may further contribute to the severity of cachexia [51].

The research on C26-induced cachexia has described an elevated expression of Serpina3n at mRNA [34] and protein [32] levels and that Serpina3n is associated with muscle mass [32]. Our findings on Serpina3n upregulation support these studies and also show that of the whole muscle proteome, Serpina3n was the best predictor of cachexia at a time point when cachexia predicts survival [32]. This finding, however, does not necessarily imply that the protein is causally related to cachexia or survival. To understand whether Serpina3n is influenced by changes in muscle protein synthesis and mass, we modulated these factors by treating TB-mice with sACVR after tumour inoculation and thus only a few days before the mice were euthanised. This demonstrated that muscle Serpina3n does not seem to represent a *bona fide* marker of alterations in muscle mass because i) muscle Serpina3n also increased in the sACVR-treated mice with a rescue of muscle mass, and ii) muscle Serpina3n correlated with the loss of BM only when the sACVR-treated mice were excluded from the analysis. Instead of being a direct regulator of muscle mass, increased Serpina3n may have various other effects. For instance, it correlated with spleen mass and serum IL-6, another strongly altered responses in cancer, showing that Serpina3n may reflect the inflammatory state in various tissues. The C26-induced splenomegaly originated from erythrocyte engulfment and white blood cell proliferation, both of which remained unaffected by sACVR administered only after tumour cell injection. This finding does not support our previous findings of splenomegaly mitigation after sACVR administration also before tumour formation [4]. In addition, increased muscle Serpina3n may also have localised effects in muscle extracellular space such as evoking decreased activity of proteases and thus protecting muscle from acute injury and muscle dystrophy [52]. This putative, protective effect of Serpina3n is also of interest in C26-TB mice, in which membrane abnormalities further accentuate muscle wasting [53]. Surprisingly, Serpina3n also correlated with muscle *Nrk2* ($r = -0.72$ and -0.74 in all mice and in TB-mice, respectively, $P < 0.001$, Figure S8) suggesting a possible connection of APR to *Nrk2* downregulation.

In conclusion, our results reveal disturbed OXPHOSome and NAD⁺ metabolism as a molecular determinant of cancer cachexia, which were rescued upon ACVR ligand blockade, depending on the used administration modality. We also present evidence implicating Serpina3n as a predictor of untreated cachexia but not as a generic marker of alterations in muscle mass.

AUTHOR CONTRIBUTION

J.J.H.: Conceptualisation; Formal analysis; Funding acquisition; Investigation; Project administration; Resources; Visualisation; Writing - original draft & editing. **F.P.** Conceptualisation; Formal analysis; Funding acquisition; Investigation; Validation; Writing - review & editing. **N.P.** Formal analysis; Visualisation; Writing - review & editing. **T.N.** Conceptualisation; Investigation; Writing - review & editing. **J.H.** Investigation. **L.E.** Methodology. **J.H.L.** Formal analysis. **B.R.** Investigation. **R.S.** Formal analysis, Methodology. **M.B.** Methodology. **O.R.** Methodology; Resources. **E.P.** Formal analysis; Funding acquisition; Visualisation; Writing - review & editing. **M.L.** Data Curation, Formal analysis; Visualisation; Writing - review & editing

FUNDING

This work was supported by the Academy of Finland (grant No. 275922 to JJH and 286359 to EP) and Cancer Society of Finland to JJH. The

research leading to these results has also received funding from AIRC under IG 2018 - ID. 21963 project (FP).

ACKNOWLEDGEMENT

We acknowledge Arja Pasternack, Mika Silvennoinen, Maarit Lehti, Mervi Matero, Mervi Tulonen, Aila Ollikainen, Risto Puurtinen, Kaisa-Leena Tulla, Eliisa Kiukkanen, Minna Savela and Jouni Tukiainen, for their valuable help and technical assistance. We also wish to acknowledge Meilahti Clinical Proteomics facility, HiLIFE supported by the Biocenter Finland, and Business Finland for supporting the NADMED project. In addition, we thank Professor Anu Suomalainen Wartiovaara for providing the NADMED protocols for our use and Dr Philippe Pierre for kindly providing the anti-puromycin antibody.

CONFLICT OF INTEREST

The authors have no relevant conflict of interest to disclose.

APPENDIX A. SUPPLEMENTARY DATA

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.molmet.2020.101046>.

REFERENCES

- [1] Argiles, J.M., Stemmler, B., Lopez-Soriano, F.J., Busquets, S., 2018. Inter-tissue communication in cancer cachexia. *Nature Reviews Endocrinology* 15:9–20.
- [2] Carson, J.A., Hardee, J.P., VanderVeen, B.N., 2016. The emerging role of skeletal muscle oxidative metabolism as a biological target and cellular regulator of cancer-induced muscle wasting. *Seminars in Cell & Developmental Biology* 54:53–67.
- [3] Kazemi-Bajestani, S.M., Mazurak, V.C., Baracos, V., 2016. Computed tomography-defined muscle and fat wasting are associated with cancer clinical outcomes. *Seminars in Cell & Developmental Biology* 54:2–10.
- [4] Nissinen, T.A., Henttilä, J., Penna, F., Lampinen, A., Lautaoja, J.H., Fachada, V., et al., 2018. Treating cachexia using soluble ACVR2B improves survival, alters mTOR localization, and attenuates liver and spleen responses. *Journal of Cachexia, Sarcopenia and Muscle* 9:514–529.
- [5] Zhou, X., Wang, J.L., Lu, J., Song, Y., Kwak, K.S., Jiao, Q., et al., 2010. Reversal of cancer cachexia and muscle wasting by ActRIIB antagonism leads to prolonged survival. *Cell* 142:531–543.
- [6] Toledo, M., Busquets, S., Penna, F., Zhou, X., Marmonti, E., Betancourt, A., et al., 2016. Complete reversal of muscle wasting in experimental cancer cachexia: additive effects of activin type II receptor inhibition and beta-2 agonist. *International Journal of Cancer* 138:2021–2029.
- [7] Chen, J.L., Walton, K.L., Winbanks, C.E., Murphy, K.T., Thomson, R.E., Makanji, Y., et al., 2014. Elevated expression of activins promotes muscle wasting and cachexia. *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology* 28:1711–1723.
- [8] Loumaye, A., de Bary, M., Nachit, M., Lausé, P., Frateur, L., van Maanen, A., et al., 2015. Role of Activin A and myostatin in human cancer cachexia. *Journal of Clinical Endocrinology & Metabolism* 100:2030–2038.
- [9] McPherron, A.C., Lawler, A.M., Lee, S.J., 1997. Regulation of skeletal muscle mass in mice by a new TGF-beta superfamily member. *Nature* 387:83–90.
- [10] Hammers, D.W., Merscham-Banda, M., Hsiao, J.Y., Engst, S., Hartman, J.J., Sweeney, H.L., 2017. Supraphysiological levels of GDF11 induce striated muscle atrophy. *EMBO Molecular Medicine* 9:531–544.
- [11] Chen, J.L., Walton, K.L., Hagg, A., Colgan, T.D., Johnson, K., Qian, H., et al., 2017. Specific targeting of TGF-beta family ligands demonstrates distinct roles in the regulation of muscle mass in health and disease. *Proceedings of the National Academy of Sciences of the United States of America* 114:E5266–E5275.

- [12] Hulmi, J.J., Oliveira, B.M., Silvennoinen, M., Hoogaars, W.M., Ma, H., Pierre, P., et al., 2013. Muscle protein synthesis, mTORC1/MAPK/Hippo signaling, and capillary density are altered by blocking of myostatin and activins. *American Journal of Physiology - Endocrinology And Metabolism* 304:E41–E50.
- [13] Lee, S.J., Reed, L.A., Davies, M.V., Girgenrath, S., Goad, M.E., Tomkinson, K.N., et al., 2005. Regulation of muscle growth by multiple ligands signaling through activin type II receptors. *Proceedings of the National Academy of Sciences of the United States of America* 102:18117–18122.
- [14] Rahimov, F., King, O.D., Warsing, L.C., Powell, R.E., Emerson Jr., C.P., Kunkel, L.M., et al., 2011. Gene expression profiling of skeletal muscles treated with a soluble activin type IIB receptor. *Physiological Genomics* 43: 398–407.
- [15] Nissinen, T.A., Degerman, J., Rasanen, M., Poikonen, A.R., Koskinen, S., Mervaala, E., et al., 2016. Systemic blockade of ACVR2B ligands prevents chemotherapy-induced muscle wasting by restoring muscle protein synthesis without affecting oxidative capacity or atrogenes. *Scientific Reports* 6:32695.
- [16] Attie, K.M., Borgstein, N.G., Yang, Y., Condon, C.H., Wilson, D.M., Pearsall, A.E., et al., 2013. A single ascending-dose study of muscle regulator ace-031 in healthy volunteers. *Muscle & Nerve* 47:416–423.
- [17] Hentila, J., Nissinen, T.A., Korkmaz, A., Lensu, S., Silvennoinen, M., Pasternack, A., et al., 2019. Activin receptor ligand blocking and cancer have distinct effects on protein and redox homeostasis in skeletal muscle and liver. *Frontiers in Physiology* 9:1917.
- [18] Lautaoja, J.H., Lalowski, M., Nissinen, T.A., Hentila, J., Shi, Y., Ritvos, O., et al., 2019. Muscle and serum metabolomes are dysregulated in colon-26 tumor-bearing mice despite amelioration of cachexia with activin receptor type 2B ligand blockade. *American Journal of Physiology - Endocrinology And Metabolism* 316:E852–E865.
- [19] Pekkala, S., Keskitalo, A., Kettunen, E., Lensu, S., Nykanen, N., Kuopio, T., et al., 2019. Blocking activin receptor ligands is not sufficient to rescue cancer-associated gut microbiota-A role for gut microbial flagellin in colorectal cancer and cachexia? *Cancers* 11 <https://doi.org/10.3390/cancers11111799>.
- [20] Goodman, C.A., Mabrey, D.M., Frey, J.W., Miu, M.H., Schmidt, E.K., Pierre, P., et al., 2011. Novel insights into the regulation of skeletal muscle protein synthesis as revealed by a new nonradioactive in vivo technique. *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology* 25:1028–1039.
- [21] Schmidt, E.K., Clavarino, G., Ceppi, M., Pierre, P., 2009. SUNSET, a non-radioactive method to monitor protein synthesis. *Nature Methods* 6:275–277.
- [22] Scifo, E., Szwajda, A., Soliymani, R., Pezzini, F., Bianchi, M., Dapkunas, A., et al., 2015. Proteomic analysis of the palmitoyl protein thioesterase 1 interactome in SH-SY5Y human neuroblastoma cells. *Journal of Proteomics* 123:42–53.
- [23] Lalowski, M.M., Bjork, S., Finckenberg, P., Soliymani, R., Tarkia, M., Calza, G., et al., 2018. Characterizing the key metabolic pathways of the neonatal mouse heart using a quantitative combinatorial omics approach. *Frontiers in Physiology* 9:365.
- [24] Laakkonen, E.K., Soliymani, R., Karvinen, S., Kaprio, J., Kujala, U.M., Baumann, M., et al., 2017. Estrogenic regulation of skeletal muscle proteome: a study of premenopausal women and postmenopausal MZ cotwins discordant for hormonal therapy. *Aging Cell* 16:1276–1287.
- [25] Babicki, S., Arndt, D., Marcu, A., Liang, Y., Grant, J.R., Maciejewski, A., et al., 2016. Heatmapper: web-enabled heat mapping for all. *Nucleic Acids Research* 44:W147–W153.
- [26] Arganda-Carreras, I., Kaynig, V., Rueden, C., Eliceiri, K.W., Schindelin, J., Cardona, A., et al., 2017. Trainable Weka Segmentation: a machine learning tool for microscopy pixel classification. *Bioinformatics (Oxford, England)* 33:2424–2426.
- [27] Gali Ramamoorthy, T., Laverny, G., Schlagowski, A.I., Zoll, J., Messaddeq, N., Bornert, J.M., et al., 2015. The transcriptional coregulator PGC-1beta controls mitochondrial function and anti-oxidant defence in skeletal muscles. *Nature Communications* 6:10210.
- [28] Burri, L., Thoresen, G.H., Berge, R.K., 2010. The role of PPARalpha activation in liver and muscle. *PPAR Research*, 2010. <https://doi.org/10.1155/2010/542359>.
- [29] Kjobsted, R., Hingst, J.R., Fentz, J., Foretz, M., Sanz, M.N., Pehmoller, C., et al., 2018. AMPK in skeletal muscle function and metabolism. *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology* 32:1741–1777.
- [30] Houtkooper, R.H., Pirinen, E., Auwerx, J., 2012. Sirtuins as regulators of metabolism and healthspan. *Nature Reviews Molecular Cell Biology* 13:225–238.
- [31] Bonetto, A., Aydogdu, T., Kunzevitzky, N., Guttridge, D.C., Khuri, S., Koniaris, L.G., et al., 2011. STAT3 activation in skeletal muscle links muscle wasting and the acute phase response in cancer cachexia. *PLoS One* 6: e22538.
- [32] Gueugneau, M., d'Hose, D., Barbe, C., de Barsy, M., Lause, P., Maiter, D., et al., 2018. Increased Serpina3n release into circulation during glucocorticoid-mediated muscle atrophy. *Journal of Cachexia, Sarcopenia and Muscle* 9:929–946.
- [33] Barreto, R., Mandili, G., Witzmann, F.A., Novelli, F., Zimmers, T.A., Bonetto, A., 2016. Cancer and chemotherapy contribute to muscle loss by activating common signaling pathways. *Frontiers in Physiology* 7:472.
- [34] Shum, A.M., Fung, D.C., Corley, S.M., McGill, M.C., Bentley, N.L., Tan, T.C., et al., 2015. Cardiac and skeletal muscles show molecularly distinct responses to cancer cachexia. *Physiological Genomics* 47:588–599.
- [35] Ballaro, R., Beltra, M., De Lucia, S., Pin, F., Ranjbar, K., Hulmi, J.J., et al., 2019. Moderate exercise in mice improves cancer plus chemotherapy-induced muscle wasting and mitochondrial alterations. *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology* 33: 5482–5494.
- [36] Romanello, V., Guadagnin, E., Gomes, L., Roder, I., Sandri, C., Petersen, Y., et al., 2010. Mitochondrial fission and remodelling contributes to muscle atrophy. *The EMBO Journal* 29:1774–1785.
- [37] Kainulainen, H., Papaioannou, K.G., Silvennoinen, M., Autio, R., Saarela, J., Oliveira, B.M., et al., 2015. Myostatin/activin blocking combined with exercise reconditions skeletal muscle expression profile of mdx mice. *Molecular and Cellular Endocrinology* 399:131–142.
- [38] Hulmi, J.J., Oliveira, B.M., Silvennoinen, M., Hoogaars, W.M., Pasternack, A., Kainulainen, H., et al., 2013. Exercise restores decreased physical activity levels and increases markers of autophagy and oxidative capacity in myostatin/activin-blocked mdx mice. *American Journal of Physiology - Endocrinology And Metabolism* 305:E171–E182.
- [39] Barbe, C., Bray, F., Gueugneau, M., Devassine, S., Lause, P., Tokarski, C., et al., 2017. Comparative proteomic and transcriptomic analysis of follistatin-induced skeletal muscle hypertrophy. *Journal of Proteome Research* 16: 3477–3490.
- [40] Alyodawi, K., Vermeij, W.P., Omairi, S., Kretz, O., Hopkinson, M., Solagna, F., et al., 2019. Compression of morbidity in a progeroid mouse model through the attenuation of myostatin/activin signalling. *Journal of Cachexia, Sarcopenia and Muscle* 10:662–686.
- [41] Puolakkainen, T., Ma, H., Kainulainen, H., Pasternack, A., Rantalainen, T., Ritvos, O., et al., 2017. Treatment with soluble activin type IIB-receptor improves bone mass and strength in a mouse model of Duchenne muscular dystrophy. *BMC Musculoskeletal Disorders* 18:20, 2016-1366-3.
- [42] Magga, J., Vainio, L., Kilpio, T., Hulmi, J.J., Taponen, S., Lin, R., et al., 2019. Systemic blockade of ACVR2B ligands protects myocardium from acute ischemia-reperfusion injury. *Molecular Therapy : The Journal of the American Society of Gene Therapy* 27:600–610.
- [43] White, J.P., Baltgalvis, K.A., Puppa, M.J., Sato, S., Baynes, J.W., Carson, J.A., 2011. Muscle oxidative capacity during IL-6-dependent cancer cachexia. *American Journal of Physiology, Regulatory, Integrative and Comparative Physiology* 300:R201–R211.

- [44] Rajman, L., Chwalek, K., Sinclair, D.A., 2018. Therapeutic potential of NAD-boosting molecules: the in vivo evidence. *Cell Metabolism* 27:529–547.
- [45] Jokinen, R., Pirnes-Karhu, S., Pietilainen, K.H., Pirinen, E., 2017. Adipose tissue NAD(+)-homeostasis, sirtuins and poly(ADP-ribose) polymerases -important players in mitochondrial metabolism and metabolic health. *Redox Biology* 12:246–263.
- [46] Fletcher, R.S., Ratajczak, J., Doig, C.L., Oakey, L.A., Callingham, R., Da Silva Xavier, G., et al., 2017. Nicotinamide riboside kinases display redundancy in mediating nicotinamide mononucleotide and nicotinamide riboside metabolism in skeletal muscle cells. *Molecular Metabolism* 6:819–832.
- [47] Deloux, R., Tannous, C., Ferry, A., Li, Z., Mericskay, M., 2018. Aged nicotinamide riboside kinase 2 deficient mice present an altered response to endurance exercise training. *Frontiers in Physiology* 9:1290.
- [48] Cray, C., Zaias, J., Altman, N.H., 2009. Acute phase response in animals: a review. *Comparative Medicine* 59:517–526.
- [49] Whitham, M., Parker, B.L., Friedrichsen, M., Hingst, J.R., Hjorth, M., Hughes, W.E., et al., 2018. Extracellular vesicles provide a means for tissue crosstalk during exercise. *Cell Metabolism* 27:237–251 e4.
- [50] Falconer, J.S., Fearon, K.C., Ross, J.A., Elton, R., Wigmore, S.J., Garden, O.J., et al., 1995. Acute-phase protein response and survival duration of patients with pancreatic cancer. *Cancer* 75:2077–2082.
- [51] Stephens, N.A., Skipworth, R.J., Fearon, K.C., 2008. Cachexia, survival and the acute phase response. *Current Opinion in Supportive and Palliative Care* 2: 267–274.
- [52] Tjondrokoesoemo, A., Schips, T., Kanisicak, O., Sargent, M.A., Molkentin, J.D., 2016. Genetic overexpression of Serpina3n attenuates muscular dystrophy in mice. *Human Molecular Genetics* 25:1192–1202.
- [53] Acharyya, S., Butchbach, M.E., Sahenk, Z., Wang, H., Saji, M., Carathers, M., et al., 2005. Dystrophin glycoprotein complex dysfunction: a regulatory link between muscular dystrophy and cancer cachexia. *Cancer Cell* 8:421–432.